

RESEARCH ARTICLE

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Regulation of inflammatory arthritis by the upstream kinase mitogen activated protein kinase kinase 7 in the c-Jun N-Terminal kinase pathway

Sang-il Lee^{1,2}, David L Boyle¹, Andres Berdeja³ and Gary S Firestein^{1*}

Abstract

Introduction: The c-Jun N-terminal kinase (JNK) is a key regulator of matrix metalloproteinase (MMP) and cytokine production in rheumatoid arthritis (RA) and JNK deficiency markedly protects mice in animal models of arthritis. Cytokine-induced JNK activation is strictly dependent on the mitogen-activated protein kinase kinase 7 (MKK7) in fibroblast-like synoviocytes (FLS). Therefore, we evaluated whether targeting MKK7 using anti-sense oligonucleotides (ASO) would decrease JNK activation and severity in K/BxN serum transfer arthritis.

Methods: Three 2'-O-methoxyethyl chimeric ASOs for MKK7 and control ASO were injected intravenously in normal C57BL/6 mice. PBS, control ASO or MKK7 ASO was injected from Day -8 to Day 10 in the passive K/BxN model. Ankle histology was evaluated using a semi-quantitative scoring system. Expression of MKK7 and JNK pathways was evaluated by quantitative PCR and Western blot analysis.

Results: MKK7 ASO decreased MKK7 mRNA and protein levels in ankles by about 40% in normal mice within three days. There was no effect of control ASO on MKK7 expression and MKK7 ASO did not affect MKK3, MKK4 or MKK6. Mice injected with MKK7 ASO had significantly less severe arthritis compared with control ASO ($P < 0.01$). Histologic evidence of synovial inflammation, bone erosion and cartilage damage was reduced in MKK7 ASO-treated mice ($P < 0.01$). MKK7 deficiency decreased phospho-JNK and phospho-c-Jun in ankle extracts ($P < 0.05$), but not phospho-MKK4. Interleukin-1beta (IL-1 β), MMP3 and MMP13 gene expression in ankle joints were decreased by MKK7 ASO ($P < 0.01$).

Conclusions: MKK7 plays a critical regulatory role in the JNK pathway in a murine model of arthritis. Targeting MKK7 rather than JNK could provide site and event specificity when treating synovitis.

Keywords: C-Jun N-terminal kinase, Mitogen-activated protein kinase kinase 7, Rheumatoid arthritis, Anti-sense oligonucleotide

Introduction

Rheumatoid arthritis (RA) is one of the most common immune-mediated diseases and is characterized by synovial inflammation and joint destruction [1]. Mitogen-activated protein kinases (MAPKs) are highly activated in rheumatoid synovium and potentially contribute to inflammatory and destructive mechanisms [2,3]. The c-Jun N-terminal kinases (JNKs), which belong to the MAPK family, play important roles in cytokine

production and extracellular matrix degradation by regulating matrix metalloproteinase (MMP) in fibroblast-like synoviocytes (FLS) and animal models of RA [4,5]. Of the three JNK isoforms, JNK1 has been implicated as a pivotal regulator of synovial inflammation in murine arthritis due to its role in mast cell degranulation and macrophage migration [6,7].

JNK is activated via dual phosphorylation by two upstream MAPK kinases (MKKs), MKK4 and MKK7 [8-10]. The mice lacking MKK4 or MKK7 are embryonic lethal suggesting the two kinases are non-redundant and serve distinct functions [11]. Some studies suggest that these differences might be due to selective

* Correspondence: gfirestein@ucsd.edu

¹Division of Rheumatology, Allergy and Immunology, UCSD School of Medicine, La Jolla, CA, USA

Full list of author information is available at the end of the article

regulation by extracellular stimuli, distinct tissue distribution and different biochemical properties [10]. Thus, an alternative approach targeting the MKKs instead of JNK could suppress signaling responses that contribute to inflammatory arthritis but spare a subset of host defense or homeostasis pathways.

Our previous studies showed that MKK4 and MKK7 are expressed and phosphorylated in RA synovium and both are activated by cytokines in RA FLS [12]. Surprisingly, cytokine-induced JNK activation and MMP production are strictly dependent on MKK7 in cytokine-stimulated FLS and do not require MKK4 [13]. Therefore, we evaluated whether selective targeting of MKK7 using anti-sense oligonucleotides (ASOs) would block arthritis-associated JNK activation and decreased arthritis severity in K/BxN serum transfer arthritis. The data indicate that blockade MKK7 mimics the effect of JNK deficiency and suppresses inflammatory arthritis.

Materials and methods

Oligonucleotides

A series of uniform chimeric 20-mer phosphorothioate oligonucleotides containing 2'-O-methoxyethyl chimeric (2'-MOE) groups at positions 1 to 5 and 15 to 20 targeted to murine MKK7 were synthesized and purified as described (kind gift of Isis Pharmaceuticals, Inc. (Carlsbad, CA, USA) [14]. Three ASOs complementary to murine MKK7 (Gen-Bank accession number AB005654) were 5'-TCTCCTGCAGCTTCTGGTCA-3', 5'-ACTTTGGTCTCTTCCTGTGA-3' and 5'-CCGTTCA-CAGTGTCTGTCGG-3'. The sequence for control ASO was 5'-CCTTCCCTGAAGGTTCCCTCC-3'.

ASO treatment in normal mice

All animal protocols received prior approval by the institutional review board. Pathogen-free male C57BL/6 mice were purchased from The Jackson Laboratory (Bar Harbor, ME, USA) and MKK7 or control ASOs were administered to mice based upon body weight by intravenous injection (25 and 50 mg/kg). Three days after injection of ASOs, mice were sacrificed and various tissues evaluated for MKK7 gene expression.

K/BxN serum transfer arthritis and ASO treatment

To induce K/BxN serum transfer arthritis [15], serum samples were pooled from arthritic adult K/BxN mice and injected intraperitoneally (IP) as previously described [16]. C57BL/6 mice received PBS, MKK7 ASOs or control ASO (50 mg/kg) i.v. twice a week beginning on Day -8 and then administered 100 μ l of K/BxN serum on Day 0. Clinical arthritis scores were evaluated using a scale of 0 to 4 for each paw for a total score of 16. Ankle thickness was measured with a caliper placed across the ankle joint at the widest point.

Histopathologic assessment was performed using a semi-quantitative scoring system as previously described [6], including synovial inflammation, bone erosion and cartilage damage.

Quantitative real time PCR

Ankle joints were collected at study termination, dissected to remove extra-articular tissue and snap-frozen in liquid nitrogen. The specimens were pulverized and total RNA was isolated using Rneasy[®] Lipid Tissue kit per manufacturer's protocol (Qiagen, Valencia, CA, USA). MKK7, IL-1 β , MMP3 and MMP13 expressions were measured by quantitative real-time PCR as previously described [17]. The threshold cycle (Ct) values were normalized to hypoxanthine-guanine phosphoribosyl transferase (HPRT) or glyceraldehyde-3-phosphate dehydrogenase (GAPDH) expression.

Western blot analysis

Snap-frozen joints were pulverized and homogenized at 100 mg of tissue per 0.5 ml of lysis buffer. Western blot analysis was then performed as described previously [18]. Anti-MKK3, anti-MKK6 and anti-GAPDH antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Anti-MKK4, anti-MKK7, anti-phospho-MKK4, anti-JNK, anti-phospho-JNK, anti-c-Jun and anti-phospho-c-Jun antibodies were purchased from Cell Signaling Technology (Danvers, MA, USA). Immunoreactive protein was detected with Immun-Star Western C kit (Bio-Rad, Hercules, CA, USA) using VersaDoc MP4000 imaging system (Bio-Rad). Densitometry analysis was carried out with Quantity One 1-D analysis software (Bio-Rad).

Statistical analysis

Data are expressed as mean \pm SE. Arthritis scores and change of ankle thickness among PBS, control ASO or MKK7 ASO-injected groups were analyzed by one-way ANOVA and Tukey's *post-hoc* test. Comparisons between control ASO and MKK7 ASO-injected groups were analyzed by two-tailed Student's *t*-test. In all tests, *P*-value < 0.05 was considered statistically significant.

Results

MKK7 knockdown by ASO in normal C57BL/6 mice

Three different 2'-MOE chimeric ASOs (MKK7 ASO-1, -2, -3) targeting distinct regions of the MKK7 gene or control ASO were injected i.v. into normal C57BL/6 mice. Three days later ankle joints were harvested and assayed for MKK7 mRNA. As shown in Figure 1A, MKK7 mRNA levels were reduced in a dose-dependent manner (Figure 1A), with greatest inhibition at a dose of 50 mg/kg of MKK7 ASO-2 compared with control ASO (*P* < 0.05, *n* = 3 mice for each group). Control ASO had no effect. MKK7 protein levels in the ankle were also analyzed by

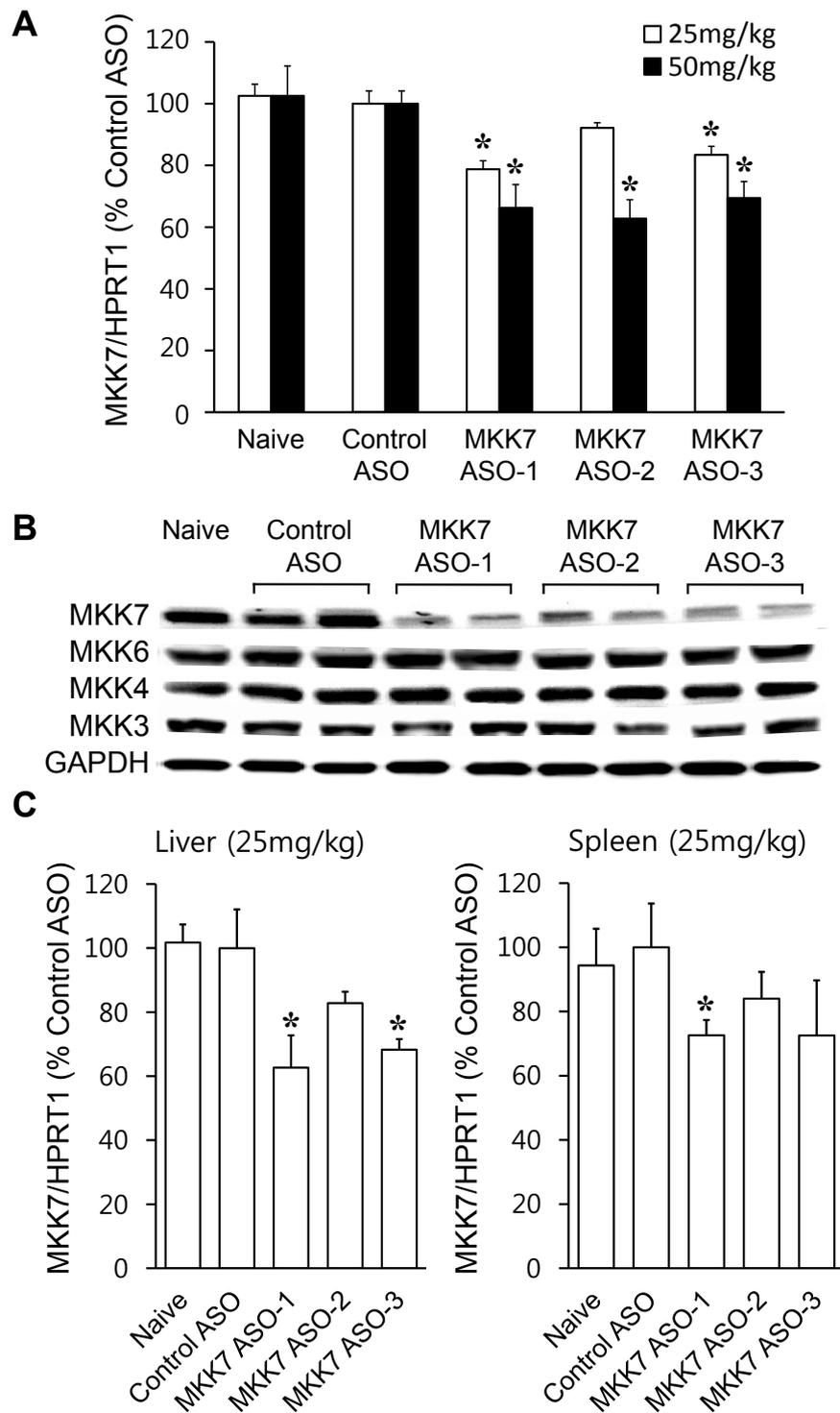


Figure 1 MKK7 ASOs selectively reduce MKK7 mRNA and protein levels in normal C57BL/6 mice. Three 2'-O-Methoxyethyl chimeric ASOs for MKK7 or control ASO were injected i.v. at the indicated doses and ankle joints, liver, and spleen were collected three days after injection of ASO. **(A)** Total mRNA was prepared from ankle joints and MKK7 mRNA expression was analyzed by quantitative PCR (Values = Mean \pm SE, $n = 3$ mice for each group, * $P < 0.05$ vs. control ASO). **(B)** Western blot analysis of proteins in ankle lysates derived from naive and control or MKK7 ASO (50 mg/kg) injected mice. MKK7 ASOs decreased MKK7 protein levels but did not decrease MKK3, MKK4, and MKK6 levels. **(C)** MKK7 ASOs decreased MKK7 mRNA levels in liver and spleen (Values = Mean \pm SE, $n = 3$ mice for each group, * $P < 0.05$ vs. control ASO).

Western blot analysis. Figure 1B shows that MKK7 protein levels were also decreased by MKK7 ASO, but MKK3, MKK4 and MKK6 levels were not affected. MKK7 mRNA was also decreased in liver and spleen by up to 37% at a dose of 25 mg/kg of MKK7 ASO-1 (Figure 1C; $P < 0.05$, $n = 3$ mice for each group) and maximally decreased in liver by up to 45% at a dose of 50 mg/kg of MKK7 ASO-1 (data not shown; $P < 0.05$).

Effect of MKK7 ASO on K/BxN serum transfer arthritis

MKK7 ASO-2 (hereafter called "MKK7 ASO") was selected for further *in vivo* experiments in passive K/BxN arthritis. C57BL/6 mice injected i.v. with PBS, MKK7 ASO or control ASO (50 mg/kg) twice a week beginning Day -8 and then administered K/BxN serum on Day 0. Mice injected with MKK7 ASO had less severe arthritis from Day 4 to Day 10 compared with control ASO (Figure 2). The peak clinical scores were 11.1 ± 0.2 in control ASO, 4.9 ± 1.0 in MKK7 ASO ($P < 0.01$) and the peak change in ankle diameter was 0.59 ± 0.06 mm in control ASO and 0.22 ± 0.06 mm in MKK7 ASO ($P < 0.01$).

Effect of MKK7 ASO on histopathology

Histopathologic analysis was performed on ankle joints obtained on Day 10 after K/BxN serum administration. Consistent with the decreased clinical arthritis, MKK7 ASO

suppressed synovial inflammation, bone erosions and cartilage destruction compared with control ASO (see Figure 3).

Effect of MKK7 ASO on MKK4, JNK, and c-Jun phosphorylation

Cytokine-induced JNK activation is dependent on MKK7 in cultured FLS and does not require MKK4. To determine the effect of selective MKK7 deficiency on JNK signaling *in vivo*, the ankle joints were evaluated by Western blot analysis to determine the phosphorylation state of MKK4, JNK and c-Jun. Consistent with the reduction of MKK7 protein level (42.2%, $P < 0.05$), MKK7 deficiency decreased GAPDH-normalized phospho-JNK by 67% and phospho-c-Jun by 62% compared with control ASO injected mice ($P < 0.05$, $n = 3$ for each group) (Figure 4). However, there was no significant difference of phosphorylation status of MKK4 between MKK7 ASO and control ASO injected groups. Similar results were obtained if the phospho-MKK4 and phospho-JNK were normalized to MKK4 and JNK, respectively. The c-Jun protein levels were higher in the control ASO-treated mice compared with MKK7 treatment due to increased local cytokine production, such as IL-1 β (see below). Thus, normalization to GAPDH provides a more reliable assessment of total phospho-c-Jun in the tissue.

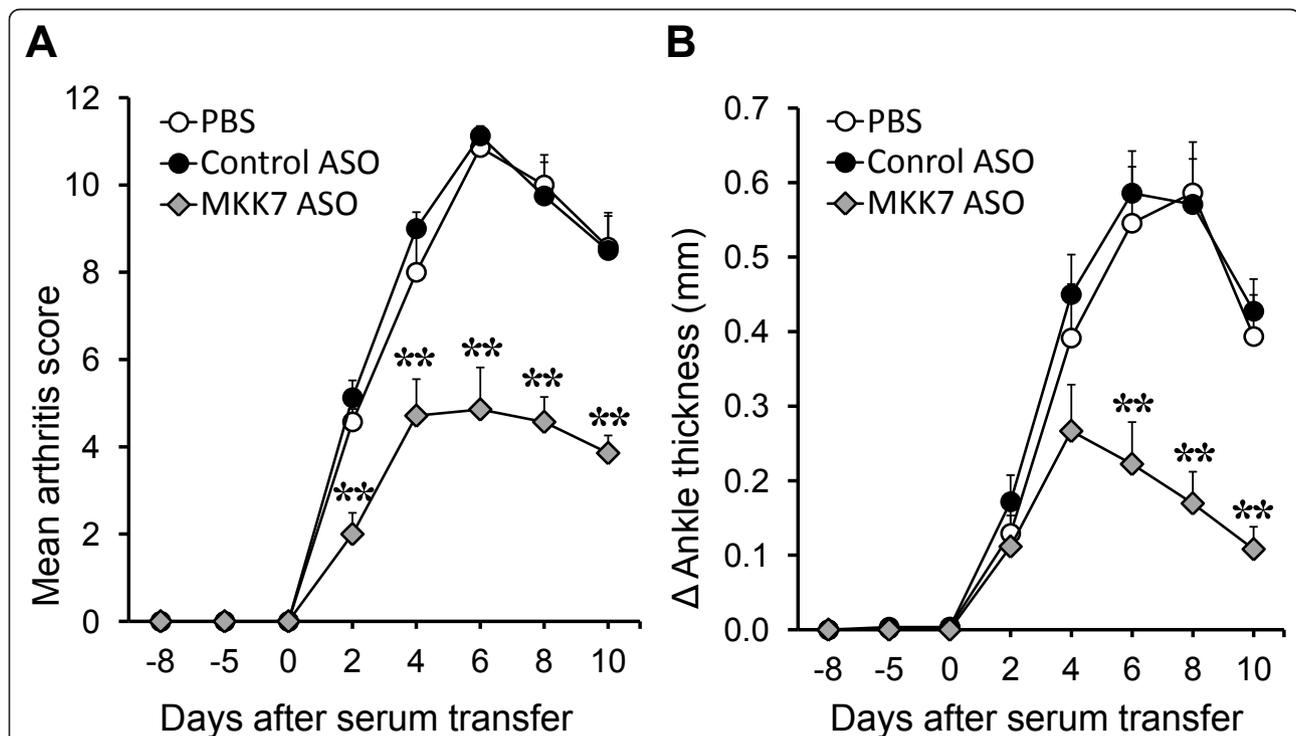
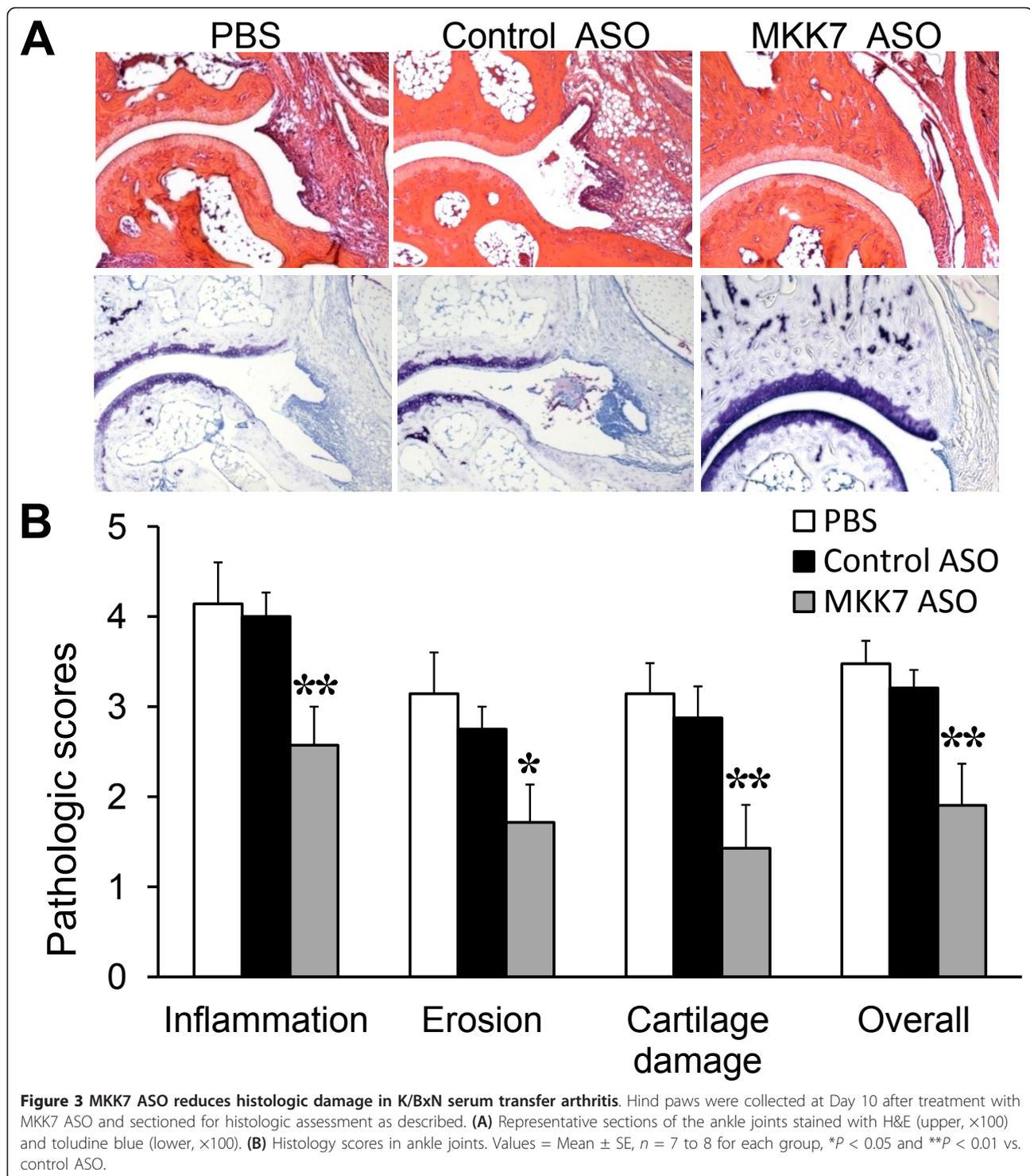


Figure 2 MKK7 ASO suppresses the arthritis severity in K/BxN serum transfer arthritis. K/BxN serum was injected on Day 0 and PBS, control or MKK7 ASO (50 mg/kg) were injected i.v. twice a week from Day -8. Arthritis severity was determined with semi-quantitative clinical scoring and change in ankle thickness. Values = Mean \pm SE, $n = 7$ to 8 mice for each group, * $P < 0.05$, ** $P < 0.01$ vs. control ASO.



Regulation of IL-1 β and MMP expression by MKK7 deficiency

The JNK pathway regulates MMP gene expression. Consistent with the reduction phospho-JNK and phospho-c-Jun in ankle joints, MMP3 and MMP13 expression were significantly decreased in the mice injected with MKK7

ASO compared with control ASO (77.9%, and 72.6%, respectively, $P < 0.05$) (Figure 5). Of interest, IL-1 β expression was also decreased. These data suggest that MKK7 plays a key role in regulating the JNK pathway, including transcription of inflammatory cytokines and proteases involved in joint damage.

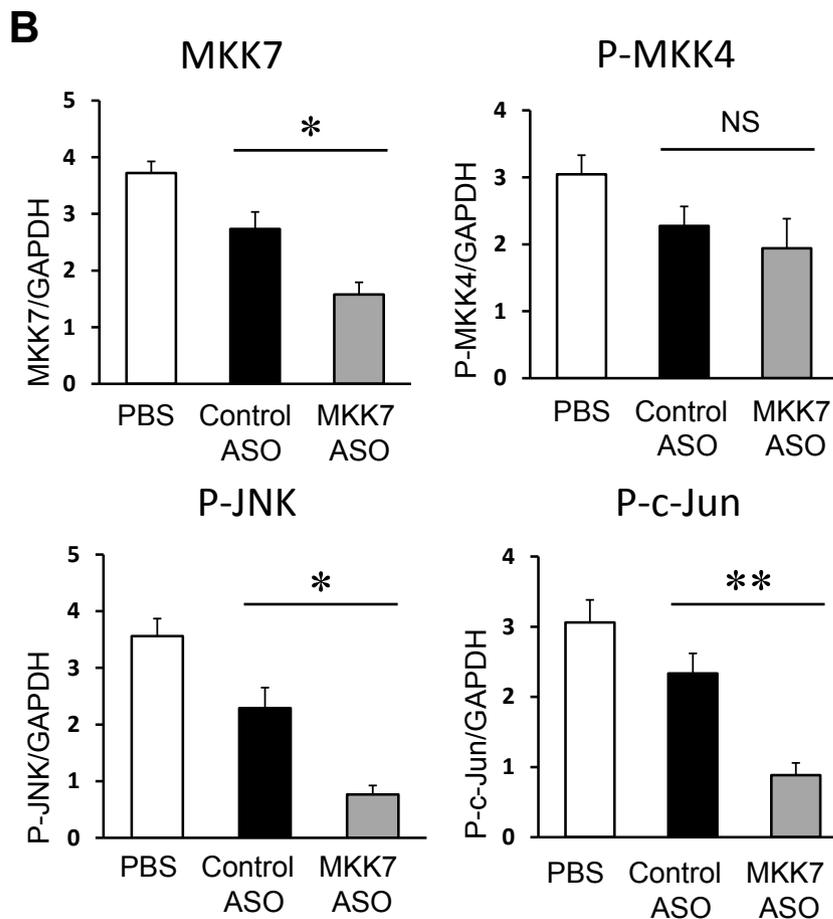
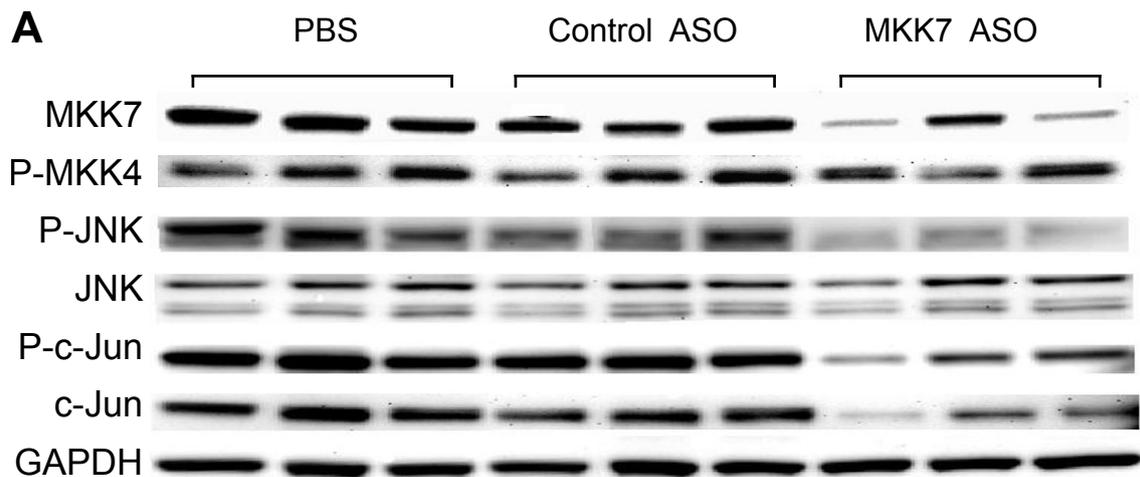
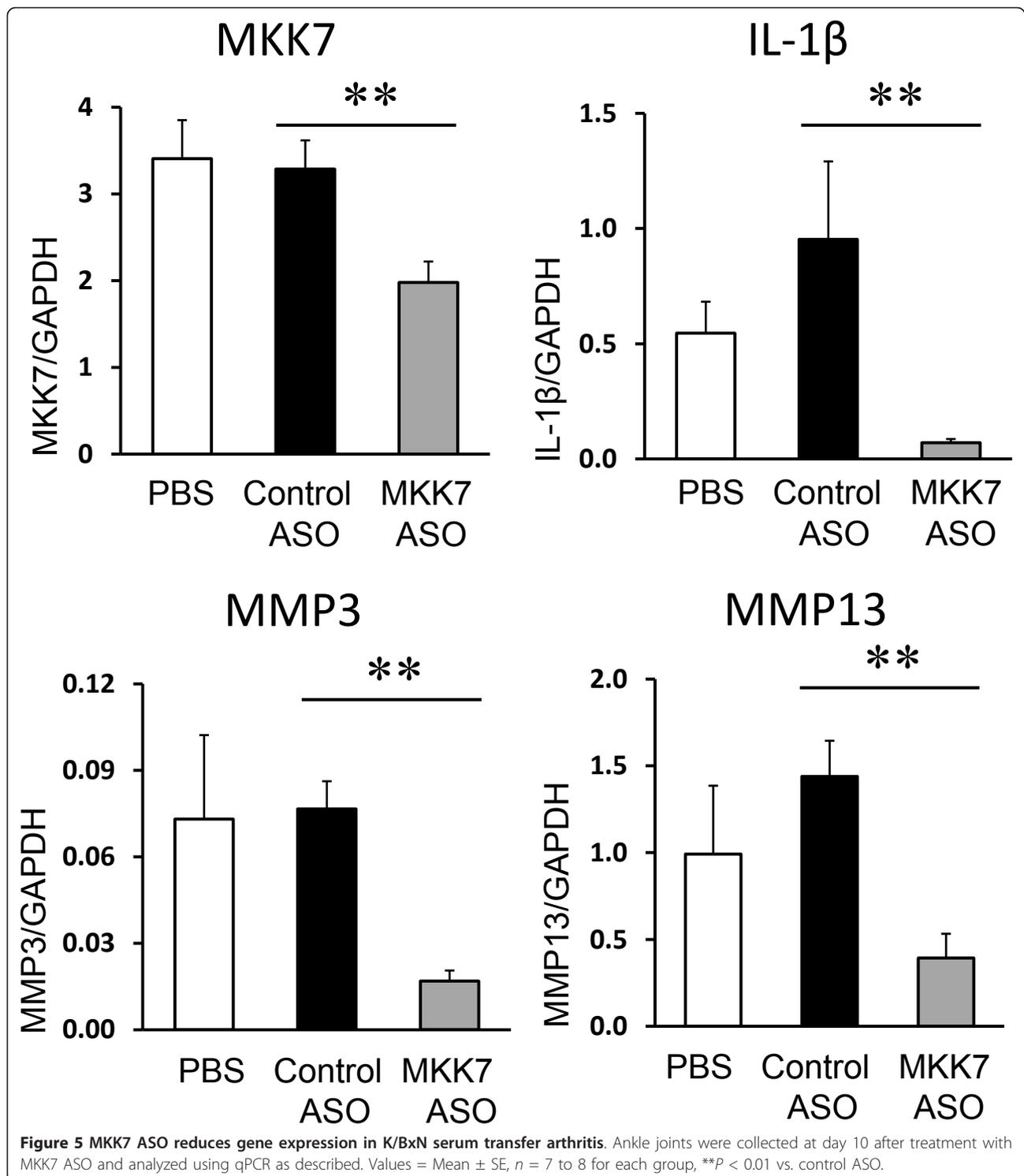


Figure 4 MKK7 ASO reduces kinase phosphorylation in K/BxN serum transfer arthritis. Ankle joints were collected at Day 10 after treatment with MKK7 ASO and homogenized as described. Western blot analysis was used to determine relative phosphorylation of MKK4, JNK, and c-Jun. **(A)** Representative experiment showing total and phosphorylated proteins. **(B)** Quantitative analysis of Western blot analysis after normalizing results to GAPDH. Values = Mean \pm SE, $n = 3$ for each group, * $P < 0.05$ vs. control ASO.



Discussion

Proinflammatory cytokines and MMPs promote synovial inflammation and facilitate cartilage and bone destruction in RA [5]. The MAPKs (ERK, JNK and p38) contribute by phosphorylating key transcription factors, such as activator protein-1 (AP-1), that are required for gene

transcription. JNK, in particular, plays a pivotal role in cytokine-mediated AP-1 induction and MMP gene expression in FLS [4,19]. Three isoforms of JNK have been characterized, namely JNK1, 2 and 3. JNK1 and 2 are ubiquitous while JNK3 is primarily restricted to neurologic tissue [20]. JNK2 deficiency has only modest

effects in pre-clinical models of arthritis, but JNK1 deficiency attenuates synovitis and joint destruction in murine antigen-induced arthritis and passive K/BxN serum transfer arthritis [6,7,21]. JNK1 also contributes to osteoclast differentiation, since JNK1-deficient osteoclast progenitors do not mature into bone-resorbing osteoclasts [22]. These data suggest that JNK participates in the synovial inflammation and joint destruction of RA and could potentially be targeted in diseases like RA.

While JNKs are attractive targets, they regulate in many normal cell functions, especially in matrix remodeling and host defense [23,24]. Thus, blocking all JNK activity, or even all JNK1 activity, could affect host defense or matrix homeostasis. As an alternative strategy, targeting an individual upstream kinase like MKK4 or MKK7 could permit some normal JNK functions while interfering with a subset that is pathogenic in synovitis. MKK4 and MKK7, two JNK upstream kinases, exhibit some different properties although they can synergistically activate JNKs [10]. TNF and IL-1 mainly activate MKK7 in murine embryonic fibroblasts, while ultraviolet radiation, anisomycin, heat and osmotic shock activate both MKK4 and MKK7 [8,9,25]. These data suggest that MKK4 and MKK7 contribute separately to the activation of JNKs in response to environmental stress or inflammatory cytokines.

We previously showed that MKK7, but not MKK4, is required for IL-1-induced JNK phosphorylation and AP-1-driven MMP expression [13]. Nevertheless, MKK4 is a component of the JNK signal complex and is also readily phosphorylated in FLS [12]. Mice lacking Gadd45 β , which serves as an endogenous inhibitor of MKK7, have enhanced JNK activity and disease severity in the passive K/BxN model [16]. These data suggest that selective MKK7 blockade could suppress arthritis and potentially decrease adverse effects by permitting non-pathogenic MKK4-mediated JNK activation. However, there is no direct evidence that MKK7 inhibition would be beneficial in synovitis. Our initial plans to focus on Gadd45 β were complicated by the recent observation that Gadd45 β deficiency unexpectedly exacerbates disease severity in collagen-induced arthritis [26].

We, therefore, focused on genetic approaches that circumvent the embryonic lethality of MKK7 deficiency. Several small interfering RNA (siRNA) methods were tested because others have reported success [27], but we were unable to consistently knockdown endogenous MKK7 expression (data not shown). Chemically modified ASOs were then tested for applications in animal models of RA because of their nuclease-resistant capacity, potency and long half-life [28-30]. Free ASOs are considerably smaller than siRNA-delivery agent complexes and enter many cells types via pinocytosis and

phagocytosis, whereas larger siRNA complexes primarily enter macrophages and neutrophils by phagocytosis [31,32]. Thus, we used single strand, 2'-O-methoxyethyl-ribose modified chimeric ASOs to investigate the effect of MKK7 deficiency in mice. Selectivity was confirmed with MKK7 ASOs, which decreased MKK7 mRNA and protein expression but not MKK3, MKK4 or MKK6.

The ASO studies showed that selective MKK7 deficiency significantly reduced arthritis severity and joint destruction compared with control ASO-injected group even though MKK7 was only partially depleted. Downstream events were consistent with previous *in vitro* studies by demonstrating reduced phosphorylation of JNK and c-Jun in the inflamed joints of MKK7 ASO-treated mice. Decreased joint damage in mice treated with MKK7 ASOs is consistent with previous observations that MKK7 is a pivotal signaling molecule that regulates JNK and MMP expression in FLS [13].

Taken together, these results imply that MKK7 plays a pivotal role in inflammatory arthritis and that MKK7 ASO acts through the inhibition of JNK in passive K/BxN arthritis. Because JNK2 does not contribute to this model, the effect is most likely due to decreased JNK1 activation with resultant decreased mast cell activation [6]. That observation is supported by the fact that JNK activation is abolished in *mkk7*^{-/-} mast cell lines, suggesting that MKK7 is essential for JNK activation in mast cells [33].

Conclusion

MKK7 plays a critical role in JNK pathway *in vivo*, and MKK7 deficiency suppresses arthritis severity and joint destruction. Selective MKK7 inhibition represents a promising alternative approach to blocking downstream kinases directly. This strategy is consistent with recent successes targeting upstream kinases like spleen tyrosine kinase (Syk) and Janus kinase (JAK) in RA and suggests that targeting upstream kinases might be useful for RA [34]

Abbreviations

AP-1: activator protein-1; ASO: anti-sense oligonucleotides; Ct: threshold cycle; ERK: extra-cellular signal regulated kinase; FLS: fibroblast-like synoviocytes; GAPDH: glyceraldehyde-3-phosphate dehydrogenase; HPRT: hypoxanthine-guanine phosphoribosyl transferase; IL: interleukin; IP: intraperitoneally; JAK: Janus kinase; JNK: c-Jun N-Terminal Kinase; MAPK: mitogen activated protein kinase; MKK7: mitogen activated protein kinase kinase 7; MMP: matrix metalloproteinase; RA: rheumatoid arthritis; Syk: spleen tyrosine kinase.

Acknowledgements

This work was supported by a grant from the National Institutes of Health (AR047825). The authors thank Meghan Edgar and Katharyn Topolewski for expert technical assistance. We also gratefully acknowledge Drs. Bill Gardner and Richard Geary from Isis Pharmaceuticals, Inc. for advice and for providing key reagents for the project.

Author details

¹Division of Rheumatology, Allergy and Immunology, UCSD School of Medicine, La Jolla, CA, USA. ²Department of Internal Medicine and Institute of Health Science, Gyeongsang National University School of Medicine, Jinju, Republic of Korea. ³Isis Pharmaceuticals, Inc., Carlsbad, CA, USA.

Authors' contributions

Each author took part in this paper and its publication is approved by all authors. SL performed all experiments, generated the figures and wrote the draft manuscript. DB participated in its design and coordination, and helped with experiments. AB performed synthesis and purifying of antisense oligomers. GSF designed, organized and analyzed the data, and wrote the manuscript.

Competing interests

Dr. Berdeja is an employee of Isis Pharmaceuticals, Inc. There are no other competing interests.

Received: 14 November 2011 Revised: 12 February 2012

Accepted: 21 February 2012 Published: 21 February 2012

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doi:10.1186/ar3750

Cite this article as: Lee et al.: Regulation of inflammatory arthritis by the upstream kinase mitogen activated protein kinase kinase 7 in the c-Jun N-Terminal kinase pathway. *Arthritis Research & Therapy* 2012 **14**:R38.